

PII: S0040-4039(96)00653-3

A SERENDIPITOUS SYNTHESIS OF CYCLOKOJIBIOSE

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Abstract: The fully protected kojibiosyl trichloroacetimidate 5 undergoes intramolecular cyclization accompanied by expulsion of the O-benzyl protecting group upon activation by a Lewis-acid to give the symmetrical, tricyclic compound 6 which is the first member of an as yet unreported group of cyclooligosaccharides. Published by Elsevier Science Ltd

Keywords: anomeric effect, cyclooligosaccharide, internal glycosylation, kojibiose, trichloroacetimidate.

The ability of cyclic oligosaccharides consisting of D-glucopyranose residues (cyclodextrins) to form inclusion complexes with hydrophobic guest compounds made them challenging synthetic targets. Cycloglycosylation strategies developed by Ogawa's group led to the total synthesis of cyclodextrins consisting of \sin^2 and $\operatorname{eight}^3 \alpha - (1 \to 4)$ -linked D-glucose residues. While the existence of smaller rings (degree of polymerization ≤ 5) in this series was questioned on the basis of potential energy calculations, the recent successful synthesis of $\operatorname{cyclo-\alpha-(1 \to 4)-gluco-pentaose^5}$ proved that $\alpha - (1 \to 4)$ -linked small-ring cyclodextrins may, indeed, be constructed however, not without distorting the ${}^4\mathrm{C}_1$ conformation of the pyranoid rings. Other reported chemical syntheses of cyclodextrins include the preparation of $\alpha - (1 \to 6)$ -linked cyclogluco-triose, tetraose, and $\alpha - (1 \to 6)$ -linked cycloglucobiose (cyclogentiobiose).

In connection with our work on α -(1 \rightarrow 2)-linked gluco-oligosaccharides¹⁰ we report here a serendipitous synthesis of cyclo- α -(1 \rightarrow 2)-glucobiose (cyclokojibiose) which is the first member of an as yet unreported group of cyclic oligosaccharides.

The protected kojibiose derivative 3^{11} was obtained in 70 % yield by the condensation of chloride 1^{10} with alcohol 2^{12} under promotion by AgClO₄/Ag₂CO₃. Regioselective deacetylation (NH₂NH₂/AcOH, Ref 13) of 3 afforded the hemiacetal 4^{11} (94 %) which was converted to the imidate 5^{11} under standard conditions 14 (87 %). (Scheme 1.) Most surprisingly, treatment of compound 5 with BF₃ · Et₂O in CH₂Cl₂ at 0 °C under rigorously anhydrous conditions either in the presence or the absence of an unreactive alcohol completely transformed compound 5 and the

Scheme 1.

Reagents and conditions: (a) 1.3 equiv. 2, 0.2 equiv. of AgClO₄, Ag₂CO₃ (excess), CH₂Cl₂, 4A molecular sieves, 25 °C, 24 h, 70%; (b) 1.3 equiv. of NH₂NH₂· AcOH, DMF, 25 °C, 3 h, 94%; (c) CCl₃CN (excess), 1,8-diazabicyclo-[5.4.0]undec-7-ene (cat), CH₂Cl₂, 0 °C, 1 h, 87%.

tricyclic compound 6^{11} could be isolated in 70 % yield. (Scheme 2.) Removal of the O-acetyl groups of 6 (NaOMe/MeOH, 80 %) afforded compound $7.^{11}$ The structure of 6 was established as follows. The most prominent peak in the chemical ionization mass spectrum at m/z 594 [(M + NH₄)+] indicated that the losses from the parent compound 5 (MW 827) exceed that would correspond to the loss of the trichloroacetimidyl moiety [CCl₃C(NH)OH] only by 90 amu, which suggests the absence of the benzyl group. Indeed, the NMR spectra were devoid of aromatic signals and consistent with either an α -linked monosaccharide having three O-acetyl groups and no other functionalities or with a highly symmetrical, α -linked saccharidic structure. The 1 H NMR spectrum recorded at 500 MHz in CDCl₃ requires the presence of acetyl groups at O-3 and O-4. Since no acetyl group can be placed on O-2, the third acetyl group must be located at O-6. The low-field position (5.21 ppm, J=3.1 Hz) of the H-1 signal indicates an unusual environment for this part of the molecule which is further corroborated by the surprisingly high-field position (90.4 ppm) for C-1 in the 13 C NMR spectrum. Removal of the O-acetyl groups (\rightarrow 7) shifted the H-3 and H-4 signals upfield as expected, while a shift in the opposite direction was observed for the H-1 signal to 5.33 ppm.

It is of importance to note that the intramolecular glycosylation reported in this work proceeded through a benzyl-protected, secondary hydroxyl group. This contrasts to cycloglycosylations reported earlier²⁻⁹ which employed intermediates with a free hydroxyl group at

Scheme 2.

Reagents and conditions: (a) 5 equiv. of $BF_3 \cdot Et_2O$, CH_2Cl_2 , 4A molecular sieves, 0 °C, 4 h 76%; (b) NaOMe (cat), MeOH, 25 °C, 3 h, 80%.

the site of the ring formation. Alternatively, this site was protected by the highly acid-labile trityl group. 15 While intramolecular expulsion of a benzyl group by a carbocation is not unprecedented 16 to our knowledge this is the first case of an anomeric carbon atom-mediated debenzylation. We propose that activation of 5 leads to a reactive intermediate in which the anomeric carbon atom $C(11)^{17}$ is placed in close proximity to the O(22) oxygen atom of the non-reducing end residue. The intermediate achieves stabilization by covalent bond formation between C(11) and O(22) and simultaneous expulsion of the benzyl group. This group is critical for the nucleophilicity of O(22) since no cycloglycosylation could be observed with the fully acetylated congener 18 of 5.

X-ray crystallographic analysis and molecular modeling studies 19 of 6 indicate that the dioxane ring interconnecting the two monosaccharide units is in a boat conformation, the C(11)-C(12)-C(21) and the C(11)-O(22)-C(22)-C(21) dihedral angles being $+4.9^{\circ}$ and -3.8° , respectively. The glucopyranose residues adopt a slightly distorted $^{4}C_{1}$ conformation. For example, the O(15)-C(11)-C(12)-C(13) dihedral angle is 47° , which is 11° smaller than the corresponding angle in methyl α -D-glucopyranoside. 20 Molecular models also show that 6 and 7 exhibit little flexibility. Their O(15)-C(15) bond is gauche to the C(11)-O(22) bond, and the *exo*-anomeric effect 21 is integrated for the dioxane ring. For the pyranose rings the anomeric effect is also integrated, but not the *exo*-anomeric effect, the H(11)-C(11)-O(22)-C(22) torsional angle being 67° . 22 The equivalent torsional angle on the other side H(21)-C(21)-O(12)-C(12) is -67° . It is likely that the combination of the stabilizing factors around the anomeric centers is sufficient to offset the unfavorable energy contributions of the dioxane moiety and the strained pyranose rings. However, the relative contributions of these effects is uncertain at this time. 23

References and Notes

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(11) Analytical and mass spectral data were consistent with the structures presented. Selected NMR data (8, CDCl₃, 20 °C except where noted otherwise) are given below. For compounds 3-5 the individual atoms are denoted by two numbers in parentheses following the atom symbol: the first one refers to the ring as defined in the Schemes, the second one refers to the atom symbol. the first one feres to the fing as defined in the Scheines, the second one feres to the individual atom of that ring. 3 $\delta_{\rm H}$ 5.79 [d, J 7.7 Hz, H(11)], 5.09 [d, J 3.4 Hz, H(21)], $\delta_{\rm C}$ 97.8 [C(21)], 92.7 [C(11)]; 4 $\delta_{\rm C}$ 99.5 [C(11β)], 97.2 and 96.2 [C(21)], 90.4 [C(11 α)]; 5 $\delta_{\rm H}$ 6.57 [d, J 3.5 Hz, H(11)], 4.92 [d, J 3.3 Hz, H(21)], $\delta_{\rm C}$ 98.4 [C(21)], 93.4 [C(11)]; 6 $\delta_{\rm H}$ 5.60 (dd, J 2.3 6.8 Hz, J 3,4 8.8 Hz, H-3), 5.21 (d, J 1,2 3.1 Hz, H-1), 4.96 (dd, J 4,5 7.8 Hz, H-4), 4.27 (ddd, J 5,6 2.8 Hz, J 5,6; 4.8 Hz, H-5), 4.39 (dd, J 6,6; -12.3 Hz, H-6'), 4.13 (dd, H-6), 3.99 (dd, H-2), $\delta_{\rm C}$ 90.4 (C-1), 72.8, 72.0, 70.8, and 67.1 (C-2,3,4,5), 61.3 (C-6); 7 (D₂O) $\delta_{\rm H}$ 5.33 (d, J 1,2 3.2 Hz, H₂ 1.3 (d, H₂ 1.3 Hz, H₂ 1.3 (d, H₂ 2.3 Hz, H₂ 1.3 (d, H₂ 2.3 Hz, H₂ 2.3 Hz, H₂ 2.3 (d, H₂ 2.3 Hz, H₂ 2.3 Hz, H₂ 2.3 (d, H₂ 2.3 Hz, H₂ 2.3 Hz, H₂ 2.3 (d, H₂ 2.3 Hz, H₂ 2.3 (d, H₂ 2.3 Hz, H₂ 2.3 Hz, H₂ 2.3 (d, H₂ 2.3 Hz, H₂ 2.3 Hz, H₂ 2.3 (d, H₂ 2.3 Hz, H₂ 2.3 Hz, H₂ 2.3 Hz, H₂ 2.3 (d, H₂ 2.3 Hz, H₂ 2.3 H 3.3 Hz, H-1), 4.04 (d, $J_{2,3}$ 7.2 Hz, H-3), 3.93 (d, $J_{2,3}$ 7.2 Hz, H-2), 3.88 (ddd, $J_{5,6}$ 2.7 Hz, $J_{5,6}$ 5.1 Hz, H-5), 3.53 (dd, $J_{4,5}$ 9.5 Hz, H-4), 3.83 (dd, $J_{6,6'}$ -12.4 Hz, H-6), 3.80 (dd, H-6'), δ_C 91.2 (C-1), 76.4 (C-2), 75.6 (C-5), 74.4 (C-3), 68.8 (C-4), 61.0 (C-6).

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